

Original Research Article

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Prevalence and Resistance Pattern of *Staphylococcus aureus* Isolated from a Al-Hussain Teaching Medical City, Al-Zahraa Hospital, and Obstetric Teaching Hospital in Karbala Governorate, Iraq

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ABSTRACT

Staphylococcus aureus continues to be a dangerous pathogen for both community-acquired as well as hospital-associated infections. The aim of this study was to determine the prevalence and antibiotic susceptibility of *S. aureus* and Methicillin Resistant *S. aureus* (MRSA) in the environments of three hospitals in Karbala, Iraq. To study the prevalence of resistant strains of *S. aureus* isolated from the environments of three hospitals in Karbala, Iraq. A total of 200 samples were collected exudative specimens such as Pus, Wound swabs, Blood, Throat swabs, and urine obtained from cultures of specimens from patients who had been hospitalized for 48 hours. The swab samples were directly plated on Mannitol Salt and Baird Parker agar plates and incubated at 37 °C (± 2) for 18–24 h. An antibiotic susceptibility test was performed using the Clinical Laboratory Standard Institute's guidelines. A total number of 200 clinical samples were processed which included 45 Pus, 40 Wound swabs, 35 Blood, 30 Throat swabs, 50 Urine. Of the entire number of clinical samples processed, only 200 samples were found positive for bacterial growth. The prevalence of *Staphylococcus aureus* in the culture positive samples was found 44% (88). Coagulase negative staphylococci, 29.16% (7) and 62.22% (38) Gram negative bacilli. The incidence of *S. aureus* in the types of clinical samples Methicillin-resistant *S. aureus* had high prevalence among Iraqi patients which were admitted in Hospitals, which indicate their predominance in our community. Accurate and continuous surveillance of antibiotic resistance patterns among *S. aureus* strains should be considered in emergency health care centers.

Keywords

Antimicrobial susceptibility, Prevalence, Antibiotic resistance, *Staphylococcus*

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Introduction

Staphylococcus aureus has long been considered as a major pathogen of hospital infections all-around the world. It has been associated with bacteremia, infective endocarditis and various types of infections including skin and soft tissue, osteoarticular, pleuropulmonary, urinary tract and device-

related (Shrestha *et al.*, 2009; Megged, 2014; Yahaghi *et al.*, 2014). The ability of *S. aureus* to develop resistance to certain environmental conditions and also wide range of antibiotics and disinfectant agents. Bacteria have been implicated as a cause of long-term survival pathogens in the environment (Holland, 2014). Methicillin resistant *S. aureus* (MRSA), has emerged as a nosocomial pathogen of major

worldwide importance and is an increasingly frequent cause of community-acquired infections that cause significant morbidity and mortality ((Rubin *et al.*, 1999). In 1961, there were reports from the United Kingdom of *S. aureus* isolates, which resisted to methicillin, and MRSA isolates were soon recovered from other European countries, and later from Japan, Australia, and the United States (CDC, 1999).

MRSA is a resistant variant of *Staphylococcus aureus* which has evolved an ability to survive treatment with beta-lactam antibiotics which includes penicillin, methicillin and cephalosporins and to various other groups of antimicrobial agents. They are often referred to as superbugs. Most isolates remain susceptible to Glycopeptides (Vancomycin, Teicoplanin), Oxazolidinones (linezolid) Streptogramins (quinupristin-dalfopristin), and polycyclic compounds (tetracycline, tigecycline) (Moreillo *et al.*, 2005; Deresinski, 2005). The synthesis of large numbers of antibiotics over the past three decades has caused complacency about the threat of bacterial resistance. Bacteria have become resistant to antimicrobial agents as a result of chromosomal changes or the exchange of genetic material via plasmids and transposons (Rubin *et al.*, 1999). Resistance to antibiotics is a significant worldwide problem and antibiotic use is being recognized as the key selective force driving this resistance (Kreiswirth *et al.*, 1993; Sehhati *et al.*, 2009). Traditionally, MRSA was identified infrequently from patients in the community, but over the last few years reports have documented increases in community-acquired MRSA, which may suggest a changing in epidemiology (Kallen *et al.*, 2000; Ghojzadeh *et al.*, 2014).

In Iraq, several studies were carried out regarding the prevalence and antimicrobial resistance of MRSA from different clinical

materials, but little is known about these bacteria in patients with health care centre. For that this study was designed to detect the prevalence and antimicrobial resistance pattern of MRSA isolates from Iraqi patients admitted in Al-Hussain Teaching medical city, Al-Zahraa hospital, and Obstetric teaching hospital in Karbala governorate / Iraq. The aim of this research was to study the prevalence of *S. aureus* infections in the Karbala hospitals and the drug resistance pattern in the isolates with an intention to help start the appropriate empirical antibiotic treatment of patients even on the levels of primary healthcare centers

Materials and Methods

Study Design

Staphylococcus aureus strains isolated from cultures of specimens from patients who have been hospitalized for > 48 hours in Al-Hussain Teaching medical city, Al-Zahraa hospital, and Obstetric teaching hospitals in Karbala governorate / Iraq were included in the study.

Isolation and Identification of Clinical Specimens

The samples were collected exudative specimens such as Pus, Wound swabs, Blood, Throat swabs, and urine obtained from cultures of specimens from patients who had been hospitalized for 48 hours. *Staphylococcus aureus* were characterized by their morphology on Gram staining, growth characteristics and coagulase production. The study was carried out between November 2017 to December 2017.

Antimicrobial Susceptibility Testing

The antibiotic susceptibility pattern of all the confirmed *S. aureus* were determined by Kirby-Bauer disc diffusion method against the following antibiotics as per CLSI guidelines:

Octadisc (combi 68). Cefoperazone (75 mcg), Cefpodoxime (30mcg), Ceftazidime (30mcg), Cefepime (30mcg), Meropenem (10mcg), Gentamicin (10 mcg), Amikacin (30 mcg) and Moxifloxacin (5 mcg). Muller-Hinton agar used to perform all antimicrobial susceptibility tests, and the interpretation criteria were taken according to National Committee for Clinical Laboratory Standard (NCCLS).

Screening for MRSA

Methicillin resistance was screened by disc diffusion method using 30µg cefoxitin disk (Becton Dickinson). The diameter of the zone of inhibition was measured and interpretation was done in accordance with the CLSI guidelines.

An isolate was considered to be a MRSA strain if cefoxitin inhibition zone diameter was < 21 mm (CLSI, 2007). Since *Staphylococcus aureus* can be a colonizer (Madani, 2002; Moreillon and Glauser, 2005) special emphasis was laid on the clinical significance of all the isolates. This was done by correlating with Gram stained smear examination and ascertaining significance with the clinical history.

Results and Discussion

In this study, we describe the incidence of MRSA isolation in Al-Hussain Teaching medical city, Al-Zahraa hospital, and Obstetric teaching hospital in Karbala governorate / Iraq. We determined the MRSA incidence in our facility while generating our hospital antibiogram in support of our healthcare providers to more effectively treat their patients. Thus, we determined the baseline of MRSA prevalence to better inform incoming healthcare providers. This study represents the longest survey of MRSA incidence performed in Karbala governorate in Iraq.

A total number of 200 clinical samples were processed which included 45 Pus, 40 Wound swabs, 35 Blood, 30 Throat swabs, 50 Urine. Of the entire number of clinical samples processed, only 200 samples were found positive for bacterial growth. The prevalence of *Staphylococcus aureus* in the culture positive samples was found 44% (88), Coagulase negative staphylococci, and 29.16% (7) and 62.22% (38) Gram negative bacilli. The incidence of *S. aureus* in the types of clinical samples is shown in Table 1.

The prevalence of MRSA and MSSA in the clinical samples has been shown in Table 2. The prevalence of MRSA was found to be 44% (88) in the entire studied population.

S. aureus is a leading pathogen in hospital acquired infections (HAIs). The prevalence of *S. aureus* infections was next to the Gram negative bacterial infections, but on the top of Gram positive bacterial infections. However, as the isolated Gram negative bacterial pathogens were not identified to their genera or species level, the *S. aureus* infections may be considered the top leading among all the infections in the observed Bastar population. All the studied subjects were tribal and native of Bastar region only, and pyogenic and urogenital infections were found common in them. Unhygienic mode of living and least health awareness might be a cause of ease in acquiring infections.

Overall resistance patterns of MRSA isolates included in this study were consistent with those obtained by other Iraqi researchers (Mohammed, 2011; Al-Hassnawi *et al.*, 2012), where the highest resistance rates were against β -lactam antibiotics, except carbapenems. Lack of control over antibiotic use in our country is the main reason for the development of increasing resistance to different antimicrobials among our clinical isolates including MRSA.

Table.1 Prevalence of *S. aureus* in the culture positive clinical samples

Sample	Samples (n)	<i>S. aureus</i>	Coagulase Negative <i>S. aureus</i>	Gram negative bacilli
Pus	45	24(53.33%)	7 (29.16%)	38 (62.22%)
Wound swabs	40	34 (85%)	0 (0%)	7 (17.5%)
Blood	35	19 (54.28%)	11 (57.89%)	12 (34.28)
Throat swabs.	30	14 (46.66%)	4 (28.57%)	0 (0%)
Urine	50	9 (18%)	3 (33.33%)	41 (82%)

Table.2 Prevalence of MRSA and MSSA in the clinical samples

Total clinical samples (n=200)	MRSA	MSSA
	88 (44%)	112 (56%)
Pus (n=45)	13 (28.88%)	32 (71.11%)
Wound swabs(n=40)	13 (32.5%)	27 (65.5 %)
Blood(n=35)	18 (51.5)	17 (48.5%)
Throat swabs(n=30)	30 (100%)	0 (0%)
Urine(n=50)	14 (28%)	36 (72.00)

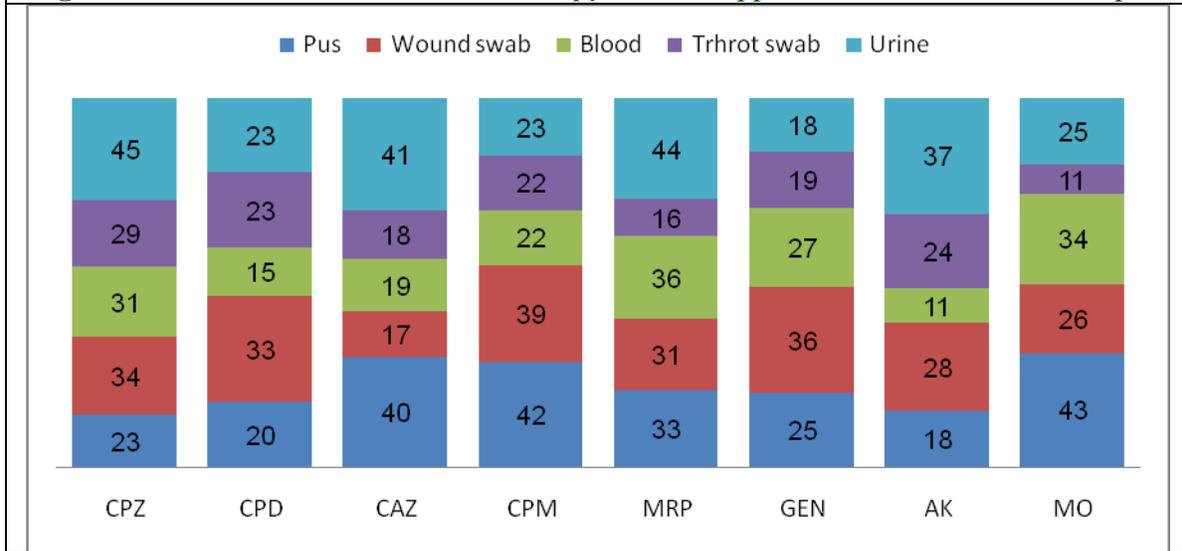
Table.3 Prevalence of MRSA and MSSA in the clinical samples

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Pus (n=45)	13 (28.88%)	32 (71.11%)
Wound swabs(n=40)	13 (32.5%)	27 (65.5 %)
Blood(n=35)	18 (51.5)	17 (48.5%)
Throat swabs(n=30)	30 (100%)	0 (0%)
Urine(n=50)	14 (28%)	36 (72.00)

Table.4 Pattern of antibiotic resistance (%)

Total clinical samples (n=200)	CPZ	CPD	CAZ	CPM	MRP	GEN	AK	MO
Pus (n=45)	23 (51.11)	20 (44.44)	40 (88.88)	42 (93.33)	33 (73.33)	25 (55.55)	18 (40)	43 (95.55)
Wound swabs (n=40)	34 (85)	33 (82.5)	17 (42.5)	39 (97.5)	31 (77.5)	36 (90)	28 (70)	26 (65)
Blood (n=35)	31 (88.57)	15 (42.85)	19 (54.28)	22 (62.85)	26 (74.28)	27 77.14	11 (31.42)	34 (37.14)
Throat swabs (n=30)	29 (96.66)	23 (76.66)	18 (60)	22 (73.33)	16 (53.33)	19 (63.33)	24 (80)	11 (36.66)
Urine (n=50)	45 (90)	23 (46)	41 (82)	23 (46)	44 (88)	18 (36)	37 (74)	25 (50)
Total (n=200)	162 (81)	114 (57)	135 (67.5)	148 (74)	150 (75)	125 (62.5)	118 (59)	139 (69.5)

Fig.1 Pattern of antibiotic resistance of *Stapylococcus* spp. isolated from clinical samples



Also, overuse of β -lactams for treating different cases in our hospitals is another reason for this high prevalence of MRSA. Emergence of antibiotic-resistant bacteria clones reflects the intensive use of antimicrobial agents (Didier *et al.*, 2011). So that there must be a scheduled rotation of β -lactams use with other antimicrobials in the study area to reduce this high antibiotic resistance among our isolates.

In a study by Alebachew *et al.*, 2012, the *S. aureus* strains of human clinical infections were sensitive to clindamycin, vancomycin, erythromycin, and kanamycin but highly resistant to penicillin. They showed that all isolates were multi-drug resistant, and one isolates was resistant to all the tested drugs. In a research in Iran, 60 % of all *S.aureus* isolates of hospital infections were resistant to methicillin.

Ekrami *et al.*, 2010, showed that the highest resistance was to ciprofloxacin (81.2 %) while Momtaz and Hafezi, 2014 found that *S. aureus* isolates from hospital infections were resistant to azithromycin (62.12 %), tetracycline (57.57 %) and erythromycin (54.54%).

Methicillin-resistant *S. aureus* had high prevalence among Iraqi patients which were admitted in Al-Hussain Teaching medical city, Al-Zahraa hospital, and Obstetric teaching hospital in Karbala governorate / Iraq, which indicate their predominance in our community. The results of the present investigation indicate that *S. aureus* might be an important causative agent of nosocomial infections in emergency centers of hospitals. Accurate and continuous surveillance of antibiotic resistance patterns among *S. aureus* strains should be considered in emergency health care centers. Effective disinfection of surfaces, beds, trolleys and surgical instruments by hospital infection control unit should be performed periodically to reduce colonization of *S. aureus* in various areas of hospital emergency wards.

References

Alebachew T, Yismaw G, Derabe A, Sisay Z. (2012). *Staphylococcus aureus* burn wound infection among patients attending yekatit 12 hospital burn units, addisababa, Ethiopia. *Ethiopian J Health Science*; 22(3): 209- 213.

- Al-Hassnawi HH, Al-Charrakh AH, Al-Khafaj JK (2012). Antibiotic resistance patterns of community acquired methicillin resistance *Staphylococcus aureus* (CA-MRSA) in Al-Hilla/ Iraq. *Kerbala Journal of Pharmaceutical Sciences* 10: 91-102.
- Centers for Disease Control and Prevention (CDC) (1999). Four Pediatric Deaths from Community-Acquired Methicillin-Resistant *Staphylococcus aureus*-Minnesota and North Dakota, 1997-1999. *Morbidity and Mortality Weekly Report (MMWR)* 1999; 48(32): 707-10.
- Clinical Laboratory Standard Institute (2007). Performance standards for antimicrobial susceptibility testing; Seventeenth informational supplement-Clinical Laboratory Standard Institute M100-S17; 27:1.
- Deresinski S. (2005). Methicillin-resistant *Staphylococcus aureus*: an evolutionary, epidemiologic, and therapeutic Odyssey. *Clin Infect Dis*; 40:562-73.
- Didier JP, Villet R, Huggler E, Lew DP, Hooper DC (2011). Impact of ciprofloxacin exposure on *Staphylococcus aureus* genomic alterations linked with emergence of rifampin resistance. *Antimicrob Agents Chemother* 55: 1848-1852.
- Ekrami A, Samarbafzadeh A, Alavi M, Kalantar E, Hamzelo F. (2010). Prevalence of methicillin resistant *Staphylococcus* species isolated from burn patients in a burn center, Ahvaz, Iran. *Jundishapur J Microbiol*; 3(2): 84-91.
- Ghojzadeh M, Naghavi-Behzad M, Nasrolah-Zadeh R, Bayat-Khajeh P, Piri R, and Mirnia K, (2014). Knowledge production status of Iranian researchers in the gastric cancer area: based on the medline database. *Asian Pac J Cancer Prev*; 15(12): 5083-8.
- Holland TL, Arnold C, Fowler VG, (2014). Clinical management of *Staphylococcus aureus* bacteremia: a review. *JAMA*; 312(13): 1330-1341.
- Kallen AJ, Driscoll TJ, Thornton S, Olson PE, Wallace MR. (2000). Increase in community-acquired methicillin-resistant *Staphylococcus aureus* at a Naval Medical Center. *Infect Control HospEpidemiol*; 21(3): 223-6.
- Madani T A. Epidemiology and Clinical features of methicillin-resistant *Staphylococcus aureus* in the University Hospital, Jeddah, Saudi Arabia. *Can J Infect Dis* 2002; 13(4): 245-250.
- Megged O. (2014). *Staphylococcus aureus* urinary tract infections in children are associated with urinary tract abnormalities and vesico-ureteral reflux. *PediatrNephrol*; 29(2): 269-272.
- Mohammed SM (2011). Use of ceftiofloxacin as indicator for detection of Methicillin Resistant *Staphylococcus aureus*. *Baghdad Science Journal* 8: 947-955.
- Momtaz H, Hafezi L. (2014). Methicillin-resistant *Staphylococcus aureus* isolated from Iranian hospitals: virulence factors and antibiotic resistance properties. *Bosn J Basic Med Sci*; 14(4): 219-226.
- Moreillon P, Que Y A, Glauser M P. (2005). *Staphylococcus aureus* (including Staphylococcal toxic shock) in Mandell, Douglas and Bennett's. Principles and practice of Infectious disease 6th ed. Churchill Livingstone; 2321-2351.
- Rubin RJ, Harrington CA, Poon A, Dietrich K, Greene JA, Moiduddin A. (1999). The economic impact of *Staphylococcus aureus* infection in New York City hospitals. *Emerg Infect Dis* 1999; 5(1): 9-17.
- Sehhati-Shafaii F, Asadollahy M, Piri R, Naghavi-Behzad M, Farzollahpour F. (2013) Prevalence and Risk Factors of Preterm Labor in Health Educational

- Centers of Northwest Iran. *Life Sci J*; 10(3): 231-236.
- Shrestha B, Pokhrel B, Mohapatra T. (2009). Study of nosocomial isolates of *Staphylococcus aureus* with special reference to methicillin resistant *S. aureus* in a tertiary care hospital in Nepal. *Nepal Med Coll J*; 11(2): 123-126.
- Yahaghi E, Imani Fooladi AA, Amin M, Mirnejad R, Nezamzade R, Amani J. (2014). Detection of Class I Integrons in *Staphylococcus aureus* isolated from Clinical Samples. *Iran Red Crescent Med J*.; 16(11): e16234.

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